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DOI: <https://doi.org/10.1016/j.febslet.2011.03.040>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-54041>

Journal Article

Accepted Version

Originally published at:

Moehle, K; Freund, A; Kubli, E; Robinson, J A (2011). NMR studies of the solution conformation of the sex peptide from *Drosophila melanogaster*. *FEBS Letters*, 585(8):1197-1202.

DOI: <https://doi.org/10.1016/j.febslet.2011.03.040>

NMR Studies of the Solution Conformation of the Sex Peptide from *Drosophila melanogaster*

Kerstin Moehle^a, Annabelle Freund^a, Eric Kubli^b and John A. Robinson^{a*}

^a Chemistry Department, University of Zurich, Winterthurerstrasse 190, 8057 Zurich, Switzerland,

^b Institute of Molecular Life Sciences, University of Zurich, Winterthurerstrasse 190, 8057 Zurich, Switzerland.

* Corresponding author

E-mail: robinson@oci.uzh.ch

Abstract

The insect sex peptide (SP) elicits a variety of biological responses upon transfer to the mated female. SP contains 36 amino acids, including a tryptophan-rich N-terminal region, a central region containing five hydroxyproline (Hyp) residues, and a C-terminal region enclosed by a disulfide bridge. The solution structure of SP, studied here using NMR spectroscopy, includes a motif WPWN that adopts a type I β -turn in the N-terminal Trp-rich region. This turn region is connected to the central Hyp-rich region, which adopts extended and/or PPII-like conformations. The C-terminal disulfide-bonded loop populates helical turns or nascent helical structure. Overall, the results reveal a rather flexible peptide that lacks a compact folded structure in solution.

Keywords:

peptide, secondary structure, conformation, sex peptide, NMR spectroscopy, fruit fly, *Drosophila*

1. Introduction

The peptide hormone known as sex peptide (SP) is produced in seminal fluid of the male fruit fly (*Drosophila melanogaster*), where it elicits a variety of responses upon transfer to the mated female. These include changes in female sexual behaviour (oviposition and receptivity), as well as food intake, egg production, Juvenile hormone (JH) synthesis, ovulation, and antimicrobial peptide synthesis [1,2]. Thus SP probably interacts with several different receptors. One of these receptors was shown recently to be a G-protein coupled receptor (GPCR) called the sex peptide receptor (SPR) or CG16752 [3], which is located in various parts of the insect neuronal and ovarian systems [4]. The interaction of SP with SPR

was shown to be responsible for eliciting oviposition and reduction of receptivity [3]. A different insect GPCR called Methuselah, which is associated with longevity, is also activated by binding to SP but the physiological relevance of this interaction remains unclear [5]. In addition, SP has been shown to elicit a systemic and epithelial innate immune response involving the production of the insect antimicrobial peptide drosocin, suggesting that SP may interact with pattern recognition receptors in the innate immune system of the insect [6,7].

The secreted mature SP contains 36 amino acids, including a tryptophan-rich N-terminal segment, a central region containing five hydroxyproline (Hyp) residues, and a C-terminal region enclosed by a disulfide bridge (Figure-1) [8,9]. The Hyp was shown to be (2*S*,4*R*)-4-hydroxyproline by comparison to a synthetic standard [9], and is consistent with the ability of all known prolyl-4-hydroxylases, including that from *D. melanogaster*, to incorporate the hydroxyl group in the *R* configuration. The biological functions of SP appear to be mediated by specific parts of the peptide [6]. Thus SPR interacts with the C-terminal disulfide-bridged loop, as this is the region needed for binding of SP to the nervous system and genital tract of females, and the same region is also essential for eliciting oviposition and reduction of receptivity [10,11]. The amino-terminal portion of SP is essential for inducing the synthesis of JH and for binding to sperm [12], whereas the Hyp-rich central region is an elicitor of the innate immune response, perhaps acting by chemical mimicry of sugar components of the bacterial cell wall [6].

In this work we investigate the conformational behavior of SP using NMR spectroscopy in water and water/trifluorethanol (TFE) solution. Information about the structure of SP in solution is of interest in understanding how this peptide hormone might interact with various different receptors in the insect nervous, ovarian and immune systems.

2. Material and Methods

2.1. Peptide synthesis

SP was synthesized on an *Applied Biosystems* peptide synthesizer ABI 433A using standard Fmoc chemistry. The following side-chain protected amino acids were used for the synthesis: Fmoc-Ala-OH, Fmoc-Asn(Trt)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gly-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Phe-OH, Fmoc-Hyp(tBu)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH and Fmoc-Trp(Boc)-OH. Fmoc deprotection was performed using 20% piperidine in NMP. Coupling reactions were carried out using HBTU/HOBt (4 eq.) in DMF and DIEA (8 eq.) in NMP. The activated amino acids (4 eq.) were coupled for 30 min. Capping was performed at the end of each cycle using a solution of HOBt (0.015 M), Ac₂O (0.5 M) and DIEA (0.125 M) in NMP.

SP was assembled on 2-chlorotrityl resin preloaded with Fmoc-Cys(Trt)-OH (0.24 mmol, 0.4 mmol/g, 600 mg). At the end of the synthesis the peptide was cleaved from the resin and deprotected in a solution of TFA:EDT:H₂O:TIS 92.5:2.5:2.5:2.5 (30 mL) with shaking for 4 h at room temp. After concentration *in vacuo*, the crude peptide was precipitated by addition of cold Et₂O (10 mL), collected by centrifugation, washed twice with Et₂O and dried *in vacuo*. The crude product was dissolved in H₂O + 1% TFA (15 mL) and the solution was stored overnight at 4°C to complete the deprotection of tryptophan residues. The crude peptide was then directly purified by reverse-phase HPLC on a preparative C₁₈ column (Zorbax Eclipse XDB-C18; 21 mm x 250 mm, 10µm, 80 Å) using a gradient of 15% to 50% acetonitrile in water (+ 0.1% TFA) over 16 min. Reduced SP was obtained as a white powder (90 mg, 20.4 µmol, 17% yield). HPLC purity >98% on an analytical C₁₈ column (Zorbax Eclipse XDB-C18; 4.6 mm x 250 mm, 5 µm, 80 Å) using a gradient of 5% to 50% acetonitrile in water (+ 0.1% TFA) over 16 min. R_t = 15.7 min. MALDI-TOF: (*m/z*) 4429.1 [M+H]⁺ (± 0.1%; M_{calc.} = 4428.1).

The reduced peptide (4.5 μmol , 20 mg) was dissolved in a solution of ammonium acetate buffer (20 ml, 50 mM, pH 8), and stirred in air at room temp. for 40 h. The peptide was purified by reverse-phase HPLC on a semi-preparative C_{18} column (Zorbax Eclipse XDB-C18; 10 mm x 250 mm, 10 μm , 80 \AA) using a gradient of 5% to 50% acetonitrile in water (+ 0.1% TFA) over 16 min. After lyophilization SP was obtained as a white solid (12 mg) in 60% yield. The product was $\geq 98\%$ pure by reverse-phase HPLC (analytical C_{18} column, Zorbax Eclipse XDB-C18; 4.6 mm x 250 mm, 5 μm , 80 \AA , using a gradient of 5% to 50% acetonitrile in water (+ 0.1% TFA) over 16 min). $R_t = 15.7$ min. MALDI-TOF: (m/z) 4427.3 $[\text{M}+\text{H}]^+$ ($\pm 0.1\%$; $M_{\text{calc.}} = 4426.3$).

Synthetic SP was assayed for oviposition and receptivity according to standard procedures [8], and was found to show full activity.

2.2 NMR spectroscopy and structure calculations

^1H NMR measurements were performed in $\text{H}_2\text{O}/\text{D}_2\text{O}$ (9:1) or pure D_2O , pH 5.0; or in a mixture (1:1) of trifluoroethanol (TFE-d_3) and $\text{H}_2\text{O}/\text{D}_2\text{O}$ (9:1), pH 5.0. Spectra were acquired on either a Bruker AV-600 or AV-700 spectrometer at 300 K. Data were processed using TOPSPIN 2.1 (Bruker) or XEASY [13]. 1D ^1H NMR spectra were also recorded at various temperatures (279 K, 286 K, 293 K, 307 K and 314 K), pH values (4, 6 and 7) and SP concentrations (0.07 mM, 0.13 mM, 0.27 mM and 0.68 mM). Water suppression was performed by presaturation. Spectral assignments were made for water and TFE/water solutions using 2D DQF-COSY, TOCSY and NOESY spectra. $^3J_{\text{HN}}$ coupling constants were determined from 1D spectra or from 2D NOESY spectra by inverse Fourier transformation of in-phase multiplets [14].

$[\text{}^{15}\text{N}, \text{}^1\text{H}]$ - and $[\text{}^{13}\text{C}, \text{}^1\text{H}]$ -HSQC spectra at natural abundance were recorded using 3 mM peptide samples in $\text{H}_2\text{O}/\text{D}_2\text{O}$ 9:1, and in pure D_2O . Acquisition data were sampled as

1024 x 128 or 1024 x 200 complex points in t2 and t1, for the ^{15}N and ^{13}C correlation spectra, respectively. The 2D data matrices were multiplied by a square-sine-bell window function and zero filled to 2k x 1k or 2k x 2k points prior to Fourier transformation. Baseline correction was applied in both dimensions. Spectra were referenced in the direct dimension to the water signal and in the ^{15}N dimension using the ^{15}N to ^1H frequency ratio of 0.101329118, while carbon referencing was directly performed on the internal TSP standard (0 ppm).

Distance restraints were obtained from NOESY spectra with a mixing time of 200 ms. Spectra were typically collected with 1024 x 256 complex data points zero-filled prior to Fourier transformation to 2048 x 1024, and transformed with a cosine-bell weighting function. The structure calculations were performed by restrained molecular dynamics in torsion angle space using DYANA [15], using only the NOE-derived distance restraints. Starting from 100 randomized conformations a bundle of 20 conformations were selected, which incur the lowest DYANA target energy function. The program MOLMOL [16] was used for structure analysis and visualization of the molecular models. The structures have been deposited in the Protein Data Base.

Time-averaged restrained MD simulations were carried out using GROMACS v4.5.3 [17] with the AMBER99 force field in explicit solvent and periodic boundary conditions. The initial structure was selected from the calculated NMR structures. The peptide was placed in a rhombic dodecahedral unit cell with a minimum distance of 1.2 nm to the box edge. TIP3P water molecules and ions to counterbalance the peptide charge were added yielding a final system of 16338 water molecules and 5 Cl^- ions. Steepest decent minimization was performed. Electrostatics were treated with particle-mesh Ewald (PME) using a short range cut-off, and a cut-off for van der Waals forces of 1.4 nm. After 20 ps equilibration with position restraints on peptide heavy atoms, a 10 ns simulation with time-averaged NOE

distance restraints (Table-S3) was performed. In all simulations the temperature was maintained close to 300 K by weak coupling to an external bath with a coupling constant of 0.1 ps. The LINCS algorithm was used to constrain all bond lengths.

3. Results and Discussion

A series of 1D ^1H NMR spectra at varying pH and peptide concentrations revealed only marginal changes in shape and dispersion of NMR signals, suggesting that the peptide does not aggregate significantly under these conditions. ^1H NMR spectra in water and in TFE/water (1:1) mixtures revealed two species in a ratio of approximately 1:3 and 1:5, respectively, which interconvert slowly on the NMR timescale, representing *cis-trans* isomers at the Trp³-Pro⁴ peptide bond. The major species was the *trans* isomer at all Xaa-Pro and Xaa-Hyp bonds, as indicated by H α -H δ NOE connectivities between the corresponding residues. When NMR spectra were recorded in D₂O, all the peptide NH groups exchanged rapidly (within the time of the NMR measurement), indicating the absence of stable intramolecular hydrogen bonded structures.

Chemical shifts. ^1H and ^{13}C chemical shift deviations (CSDs) from random coil values ($\Delta\delta = \delta^{\text{observed}} - \delta^{\text{random}}$) were examined for indications of regular secondary structure (Figure-2) [18]. The patterns expected are positive values for $\Delta\delta_{\text{H}\alpha}$ and $\Delta\delta_{\text{C}\beta}$ and negative values for $\Delta\delta_{\text{C}\alpha}$ in β -strands, whereas values of opposite sign are expected for α -helices [19]. The CSDs in the N-terminal Trp-rich region should be interpreted with caution, since they may be influenced by aromatic ring current as well as conformational effects [20], although analyses of NOEs provide evidence for a β -turn at the Trp³Pro⁴Trp⁵Asn⁶ motif (see below). In the central Hyp-rich region (the CSDs of Hyp have not been documented), the $\Delta\delta_{\text{H}\alpha}$ and $\Delta\delta_{\text{C}\alpha}$ values for non-Hyp residues clearly suggest that β -structure is favored (Figure-2). The numerous Hyp residues, however, may have a special effect on the conformation of the

peptide backbone in the central region. For example, the role of Hyp in stabilizing the triple helix in collagen is well known [21]. Also, proline residues are generally restricted to two narrow regions of dihedral space: the α -helical region at $(\phi, \psi) = (-61^\circ, -35^\circ)$ and the β -region at $(-65^\circ, +150^\circ)$ [22,23]. The extended conformation – the so-called left-handed polyproline II helical conformation (PPII) – is the dominant fold in pure polyproline peptides. The PPII conformation is an extended helical structure, with three residues per turn, which does not form intramolecular backbone hydrogen bonds [23]. While the PPII helix has a preference for proline residues, it is also found with other amino acids [23]. Furthermore, proline restricts the preceding residue to the β -region of dihedral (ϕ, ψ) space [22]. The CSDs observed for each amino acid preceding a Hyp residue are consistent with β - and/or PPII conformations. In the C-terminal region from Arg²⁰-Asn²⁷, the negative $\Delta\delta_{H\alpha}$, positive $\Delta\delta_{C\alpha}$, and negative $\Delta\delta_{CB}$ values provide support here for nascent helical secondary structure (Figure-2). From Cys²⁴ this includes several residues that form part of the disulfide-bridged loop. From Leu²⁸ to the C-terminus this trend into negative CSDs is less apparent, suggesting that nascent helical structure does not extend into the second half of the disulfide-bridged loop. Interestingly, at the end of the helical section (Arg²⁰-Asn²⁷) the motif Gly²⁹Pro³⁰Ala³¹ possesses a high propensity for β -turn formation. Indeed, NOEs are observed that support the occurrence of a nascent turn in this region of the SP backbone (see below).

All measured chemical shifts, including HA, CA, CB, HN and N (see Supporting information), were also analyzed using TALOS+, a program that exploits the large database of protein NMR data to establish accurate relations between chemical shifts and backbone torsion angles ϕ and ψ [24]. Using these data, TALOS+ predicts with a "good" score (RCI-derived order parameter S^2 from 0.69-0.77) that all non-Hyp residues from Phe¹²-Asn¹⁸ will occupy the β -region, and all residues from Trp²³-Leu²⁸ the α -region of ϕ/ψ space.

Coupling constants. Backbone $^3J_{\text{HN}\alpha}$ coupling constants ≤ 5 Hz are typical for residues in α -helices and >7 Hz for residues in regular β -structure [25]. However, most of the $^3J_{\text{HN}\alpha}$ values for SP in water are in the range 6-7 Hz (Figure 3), which is consistent with a relatively flexible backbone and/or with PPII conformations. However, a series of lower $^3J_{\text{HN}\alpha}$ values are seen in the region Arg²⁰-Asn²⁷ in water/TFE, adding further support for nascent helical structures in this region.

Nuclear Overhauser Effects. 2D-NOESY spectra revealed few long range NOEs (see below), suggesting that SP does not adopt a compact folded structure under the conditions tested (Figure-3). Average solution structures for SP were calculated with DYANA using NOE-derived distance restraints. Rather high mean rmsd values are obtained after global backbone superimposition of the final 20 structures. The observed NOEs, however, provide evidence for ordered secondary structure, or nascent structure, only in distinct regions of SP, which is supported also by analyses of CSDs and coupling constants (see above). The three regions of interest are the N-terminal Trp-rich sequence (Glu²-Lys⁸), the Hyp-rich central region (Phe¹²-Asn¹⁸), and the disulfide-bonded loop region (Lys²²-Cys³⁶). Upon superimposition of these individual regions in the final 20 structures, lower backbone rmsd values are obtained (Table-1). In addition, time-averaged NOE-derived distance restraints were applied in MD simulations in order to take account of flexibility in the SP backbone (see Supporting information, Figure S2).

In the N-terminal Trp-rich region of SP in TFE/water, sequential as well as multiple medium range NOEs (between Trp³ and Trp⁵/Asn⁶ - see Supporting information for a full list of NOE distance restraints) provide strong evidence for a well-defined type I β -turn in the WPWN motif. This turn is highly populated in the NMR structures calculated using DYANA (Figure 4). This turn occurs in the *trans*-Trp³-Pro⁴ rotamer, and is characterized by the Trp⁵ side chain stacking against the pyrrolidine ring of Pro⁴ (Figure-4B and C) and the side-chain

of Trp³. A much stronger Pro⁴ H α to Trp⁵ HN NOE than actually observed would be expected if a type II turn were formed. Evidence for a type I turn is the upfield shifted H α resonance for Pro⁴ and downfield shifted of H α in Trp⁵ (Figure 2), a combination that is typical for position 2/3 of only the type I turn [26]. On the other hand, for SP dissolved in water only two weak medium range NOEs are seen between Trp³ and Trp⁵. This suggests that a nascent rather than a stable turn is populated in this region in pure water. During the course of 10 ns time-averaged distance-restrained MD simulations (TAMD) in explicit water solvent, the Pro⁴-Trp⁵ remained in a type I turn (Figure S2).

In the hydroxyproline-rich central region (Hyp⁹-Hyp¹⁹), only intra-residue and sequential NOEs are observed in both solvents. This and the observation of strong H α (*i*)-H δ (*i*+1) and weak or absent HN(*i*)-H δ (*i*+1) NOEs, for each Xaa-Hyp pair, are consistent with extended and/or PPII conformations in this region. The same conclusion was also suggested from the CSD data, discussed above. It seems that this region may be extended and semi-rigid, as seen in the NMR structures shown in Figure 4. The TAMD simulations support the high propensity of PPII conformations in this region, although turn conformations were also observed at Ser¹⁶-Hyp¹⁷, and Hyp¹⁹-Arg²⁰ (see Supporting information).

In the region of the disulfide-bonded loop (Arg²⁰-Cys³⁶), a series of sequential HN-HN as well as medium range H α (*i*)-NH(*i*+2) and H α (*i*)-NH(*i*+3) NOEs are seen from Arg²⁰ to Asn²⁷, particularly in water/TFE mixture. These NOEs are characteristic of α -helical conformations, or a nascent helix (in water) in this region, as suggested also by the CSD data discussed above. However, the TAMD simulations (Figure-S2) suggest more flexibility may be present in this region. A reversal of the peptide chain direction is needed for disulfide ring closure at the C-terminus, which is achieved by a turn conformation at Pro³⁰-Ala³¹. A strong HN-HN NOE between Trp³²-Ala³¹, a H α -HN NOE between Trp³²-Pro³⁰, as well as the upfield shifted HN of Trp³² also support the occurrence here of turn-like structures. A type-I

β -turn conformation at Pro³⁰-Ala³¹ is also stably maintained in the TAMD trajectory (Figure S2), in agreement with the NMR data. Furthermore, long range NOEs between the side chains of Leu²⁶-Trp²³, and between Trp²³, Leu²⁸ and Trp³² indicate that these amino acid side chains are clustered into a hydrophobic patch on the same side of the macrocycle (Figure 4). Consistent with this clustering is the upfield shift of the Asn²⁷ side chain protons, which in DYANA structures come into close contact to the Trp²³ aromatic side chain. However, no evidence of regular secondary structure was observed in the final C-terminal segment between Trp³² and Cys³⁶.

The overall picture to emerge from these studies is of a flexible peptide that lacks a compact folded structure, but which nevertheless possesses elements of nascent secondary structure in distinct regions, in aqueous solution. This nascent structure becomes more pronounced in water/TFE mixtures. The N-terminal Trp-rich region contains a motif WPWN that forms type I β -turns. This turn region is connected to a central Hyp-rich region, which populates an extended and/or PPII-like conformation. Finally, towards the C-terminus and the disulfide-bonded loop, a region of nascent helical and another β -turn is found. These distinct regions of nascent secondary structure may be important for recognition of SP by its various cellular receptors.

Acknowledgements

This work was supported by the University of Zurich. Prof. Oliver Zerbe is thanked for help with measurement of heteronuclear NMR spectra.

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Figures

Figure-1. The sequence of SP. The normal single letter amino acid code is used, except for O* = *trans*-L-4-hydroxyproline. The regions associated with specific biological responses are also shown (JH = juvenile hormone).

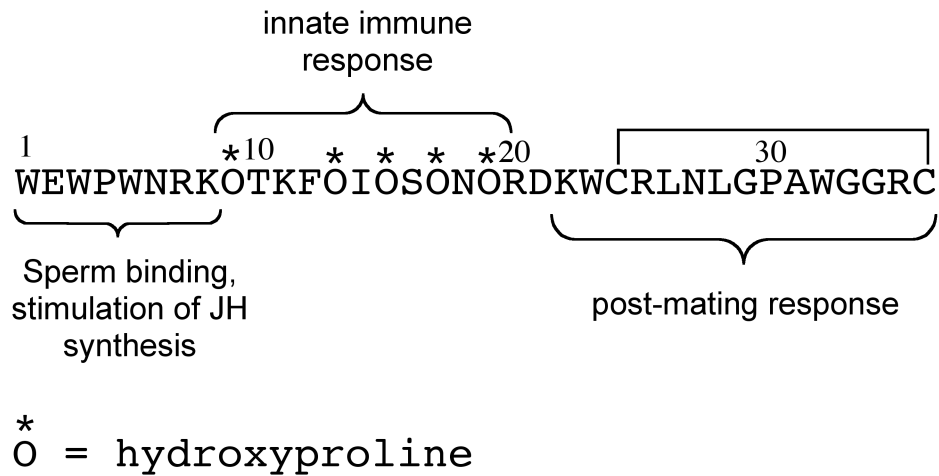


Figure-2. ^1H and ^{13}C Chemical shift deviations (CSDs) from random coil values for residues 1-36 in SP ($\Delta\delta = \delta^{\text{observed}} - \delta^{\text{random}}$) for $^1\text{H}\alpha$ (A), $^{13}\text{C}\alpha$ (B) and $^{13}\text{C}\beta$ (C) resonances in water solution. The asterisk shows the positions of hydroxyproline (Hyp) in the sequence. Values for Gly are not included.

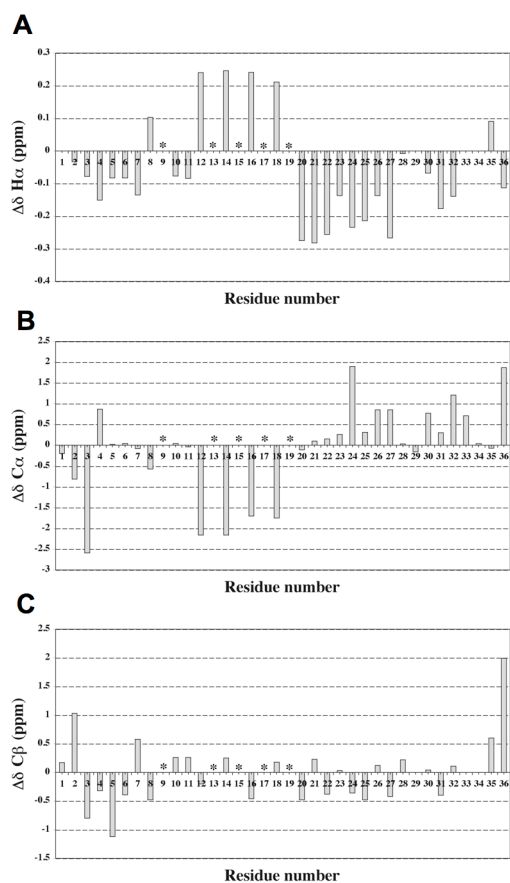


Figure-3. NOEs and $^3J_{\text{HN}\alpha}$ values observed for SP in (A) water and (B) water:TFE (1:1) solution. The letter "O" indicates hydroxyproline. NOEs shown to peptide NH groups are to the $\text{CH}_2(\delta)$ for P and O residues.

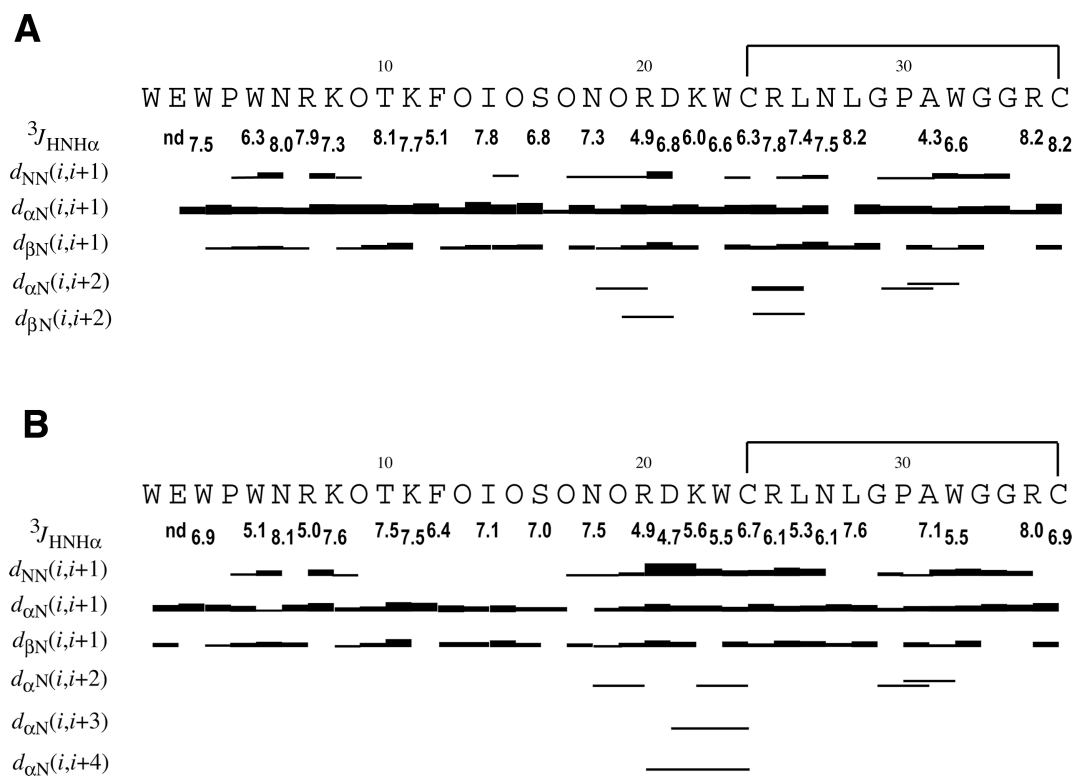


Figure-4. (A) Backbone representation and superposition of the 20 DYANA NMR structures of SP using the backbone heavy atoms of Phe¹² to Asn¹⁸ (in blue), observed in TFE/water (*left*) and water (*right*) (the N-terminal region between Trp¹ and Lys¹¹ is shown in red, the C-terminal region between Hyp¹⁹ and Cys³⁶ in green). (B) The Trp-rich N-terminal residues 1-8 in TFE/water superimposed (using the backbone heavy atoms) between Trp³ and Asn⁶ (in orange), showing the type I β -turn at WPWN. (C) Representative side chain orientation in the turn region between 3 and 6. (D) Backbone representation and superposition of SP in TFE/water between Hyp¹⁹ and Cys³⁶ (disulfide bond in yellow). (E) Ribbon representation of one solution structure of the region Hyp¹⁹ - Cys³⁶ showing the nascent helical element. The side chains of Trp²³, Leu²⁶, Leu²⁸ and Trp³² are arranged in a hydrophobic patch.

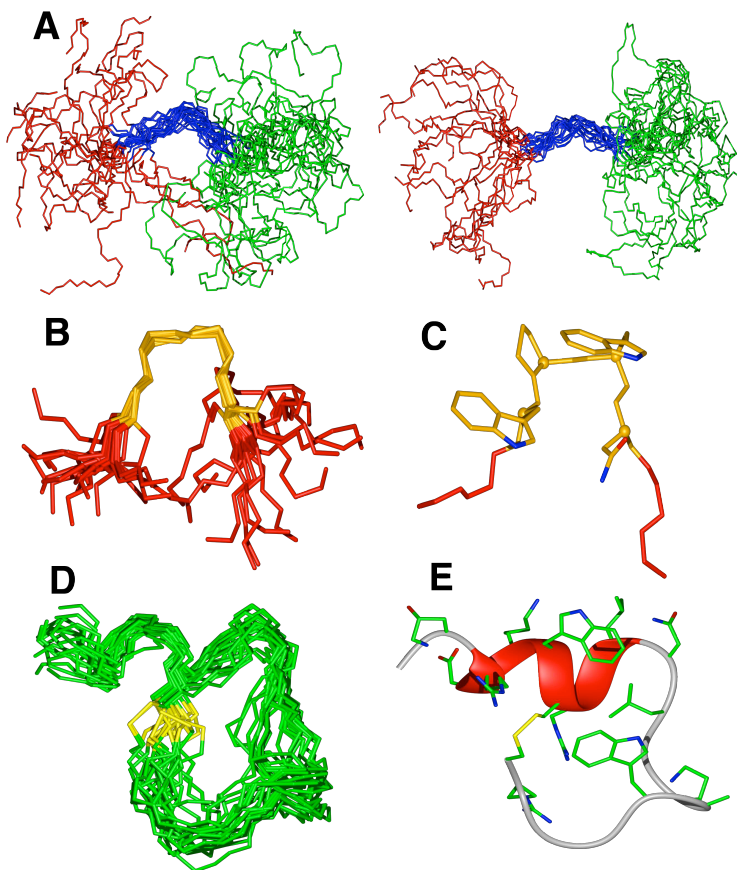


Table-1. Experimental distance restraints and statistics for the final 20 NMR structures calculated for SP in water and TFE/water.

	<i>water</i>	<i>TFE/water</i>
Number of NOE upper-distance limits	263	360
Intraresidue	101	127
Sequential	138	182
Medium- and long-range	24	51
Residual target function value (\AA^2)	2.43 ± 0.40	1.49 ± 0.11
Residual NOE violations		
Number $> 0.2 \text{ \AA}$	32	15
Maximum (\AA)	0.5	0.44
Mean rmsd values (\AA)		
<i>between Glu² and Lys⁸</i>		
backbone atoms	2.05 ± 0.46	1.62 ± 0.54
All heavy atoms	4.62 ± 0.79	3.55 ± 0.97
<i>between Phe¹² and Asn¹⁸</i>		
backbone atoms	1.96 ± 0.57	2.10 ± 0.59
All heavy atoms	3.51 ± 0.81	3.62 ± 0.84
<i>between Lys²² and Cys³⁶</i>		
backbone atoms	2.90 ± 1.31	1.97 ± 0.57
All heavy atoms	4.44 ± 1.61	3.10 ± 0.78

Supplementary data

NMR Studies of the Solution Conformation of the Sex Peptide from *Drosophila melanogaster*

Kerstin Moehle^a, Annabelle Freund^a, Eric Kubli^b and John A. Robinson^{a*}

^a Chemistry Department, University of Zurich, Winterthurerstrasse 190, 8057 Zurich, Switzerland,

^b Institute of Molecular Life Sciences, University of Zurich, Winterthurerstrasse 190, 8057 Zurich, Switzerland.

The following data are provided below:

- 1) Chemical shift assignments for SP in water (**Table S1**)
- 2) Chemical shift assignments for SP in TFE/water (**Table S2**)
- 3) A list of NOE-derived distance restraints in water (**Table S3**)
- 4) A list of NOE-derived distance restraints in TFE/water (**Table S4**)
- 5) ¹⁵N-Chemical shift deviations from random coil (**Figure S1**)
- 6) MD simulation of SP with time-averaged NOE-derived distance restraints: ϕ/ψ backbone torsion angles (**Figure-S2**).

Table S1 ^1H , ^{13}C and ^{15}N chemical shifts (ppm) of SP in water (pH 5.0, 300 K).

1	Trp	CA	57.300
1	Trp	HA	4.168
1	Trp	CB	29.770
1	Trp	HB2	3.121
1	Trp	HB3	3.198
1	Trp	NE1	129.811
1	Trp	HD1	6.921
1	Trp	HE3	7.380
1	Trp	HE1	9.672
1	Trp	HZ3	6.730
1	Trp	HZ2	7.358
1	Trp	HH2	7.015
2	Glu	N	123.180
2	Glu	HN	8.088
2	Glu	CA	55.790
2	Glu	HA	4.259
2	Glu	CB	30.930
2	Glu	HB2	1.655
2	Glu	HB3	1.817
2	Glu	HG2	2.036
2	Glu	HG3	2.086
3	Trp	N	123.710
3	Trp	HN	8.143
3	Trp	CA	54.910
3	Trp	HA	4.614
3	Trp	CB	28.800
3	Trp	QB	2.573
3	Trp	NE1	129.169
3	Trp	HD1	7.187
3	Trp	HE3	7.643
3	Trp	HE1	10.176
3	Trp	HZ3	7.250
3	Trp	HZ2	7.507
3	Trp	HH2	7.260
4	Pro	CA	64.170
4	Pro	HA	4.290
4	Pro	CB	31.780
4	Pro	HB2	1.842
4	Pro	HB3	2.207
4	Pro	HG2	1.911
4	Pro	HG3	1.956
4	Pro	HD2	3.390
4	Pro	HD3	3.801
5	Trp	N	117.357
5	Trp	HN	7.371
5	Trp	CA	57.520
5	Trp	HA	4.624
5	Trp	CB	28.480
5	Trp	QB	3.309
5	Trp	NE1	130.395

5	Trp	HD1	7.179
5	Trp	HE3	7.563
5	Trp	HE1	10.137
5	Trp	HZ3	7.169
5	Trp	HZ2	7.469
5	Trp	HH2	7.206
6	Asn	N	119.590
6	Asn	HN	7.959
6	Asn	CA	53.140
6	Asn	HA	4.671
6	Asn	CB	38.510
6	Asn	QB	2.590
6	Asn	ND2	111.560
6	Asn	HD21	7.570
6	Asn	HD22	6.875
7	Arg	N	120.540
7	Arg	HN	7.806
7	Arg	CA	55.920
7	Arg	HA	4.246
7	Arg	CB	31.480
7	Arg	HB2	1.658
7	Arg	HB3	1.806
7	Arg	HG2	1.515
7	Arg	HG3	1.565
7	Arg	QD	3.073
8	Lys	N	123.810
8	Lys	HN	8.291
8	Lys	CA	55.630
8	Lys	HA	4.477
8	Lys	CB	32.620
8	Lys	HB2	1.665
8	Lys	HB3	1.765
8	Lys	QG	1.401
8	Lys	QD	1.657
8	Lys	QE	2.978
9	Hyp	CA	61.960
9	Hyp	HA	4.582
9	Hyp	CB	39.920
9	Hyp	HB2	1.964
9	Hyp	HB3	2.319
9	Hyp	CG	72.660
9	Hyp	HG2	4.555
9	Hyp	HD2	3.689
9	Hyp	HD3	3.766
10	Thr	N	118.840
10	Thr	HN	8.359
10	Thr	CA	61.840
10	Thr	HA	4.273
10	Thr	CB	70.060
10	Thr	HB	4.124
10	Thr	QG2	1.137
10	Thr	CG2	21.680

11	Lys	N	123.510
11	Lys	HN	8.171
11	Lys	CA	56.160
11	Lys	HA	4.276
11	Lys	CB	33.360
11	Lys	HB2	1.616
11	Lys	HB3	1.690
11	Lys	HG2	1.243
11	Lys	HG3	1.304
11	Lys	QD	1.626
11	Lys	QE	2.946
12	Phe	N	122.070
12	Phe	HN	8.226
12	Phe	CA	55.540
12	Phe	HA	4.902
12	Phe	CB	39.400
12	Phe	HB2	2.893
12	Phe	HB3	3.141
12	Phe	QD	7.294
12	Phe	QE	7.352
13	Hyp	CA	61.960
13	Hyp	HA	4.573
13	Hyp	CB	39.870
13	Hyp	HB2	2.014
13	Hyp	HB3	2.296
13	Hyp	CG	72.660
13	Hyp	HG2	4.571
13	Hyp	HD2	3.700
13	Hyp	HD3	3.820
14	Ile	N	123.240
14	Ile	HN	8.399
14	Ile	CA	58.940
14	Ile	HA	4.470
14	Ile	CB	39.050
14	Ile	HB	1.880
14	Ile	QG2	0.993
14	Ile	CG1	17.300
14	Ile	HG12	1.229
14	Ile	HG13	1.575
14	Ile	QD1	0.911
14	Ile	CD1	13.100
15	Hyp	CA	61.960
15	Hyp	HA	4.583
15	Hyp	CB	39.920
15	Hyp	HB2	2.040
15	Hyp	HB3	2.341
15	Hyp	CG	72.660
15	Hyp	HG2	4.585
15	Hyp	HD2	3.823
15	Hyp	HD3	3.936
16	Ser	N	118.680
16	Ser	HN	8.583

16	Ser	CA	56.600
16	Ser	HA	4.729
16	Ser	CB	63.340
16	Ser	QB	3.843
17	Hyp	CA	61.920
17	Hyp	HA	4.570
17	Hyp	CB	39.920
17	Hyp	HB2	2.023
17	Hyp	HB3	2.316
17	Hyp	CG	72.660
17	Hyp	HG2	4.570
17	Hyp	QD	3.831
18	Asn	N	121.030
18	Asn	HN	8.762
18	Asn	CA	51.350
18	Asn	HA	4.962
18	Asn	CB	39.080
18	Asn	HB2	2.762
18	Asn	HB3	2.896
18	Asn	ND2	113.270
18	Asn	HD21	7.700
18	Asn	HD22	6.976
19	Hyp	CA	63.060
19	Hyp	HA	4.515
19	Hyp	CB	40.070
19	Hyp	HB2	2.095
19	Hyp	HB3	2.396
19	Hyp	HG2	4.612
19	Hyp	QD	3.911
20	Arg	N	118.840
20	Arg	HN	8.356
20	Arg	CA	55.890
20	Arg	HA	4.111
20	Arg	CB	30.420
20	Arg	HB2	1.725
20	Arg	HB3	1.819
20	Arg	QG	1.617
21	Asp	N	119.900
21	Asp	HN	8.047
21	Asp	CA	54.300
21	Asp	HA	4.497
21	Asp	CB	41.330
21	Asp	QB	2.675
22	Lys	N	120.130
22	Lys	HN	7.992
22	Lys	CA	56.350
22	Lys	HA	4.103
22	Lys	CB	32.720
22	Lys	QB	1.700
22	Lys	QG	1.238
22	Lys	QD	1.575
22	Lys	QE	2.864

23	Trp	N	120.070
23	Trp	HN	7.995
23	Trp	CA	57.760
23	Trp	HA	4.554
23	Trp	CB	29.630
23	Trp	HB2	3.260
23	Trp	HB3	3.334
23	Trp	NE1	129.405
23	Trp	HD1	7.240
23	Trp	HE3	7.554
23	Trp	HE1	10.149
23	Trp	HZ3	7.113
24	Cys	N	118.950
24	Cys	HN	8.268
24	Cys	CA	57.300
24	Cys	HA	4.460
24	Cys	CB	40.740
24	Cys	QB	3.039
25	Arg	N	121.070
25	Arg	HN	8.165
25	Arg	CA	56.310
25	Arg	HA	4.164
25	Arg	CB	30.420
25	Arg	HB2	1.701
25	Arg	HB3	1.799
25	Arg	CG	29.250
25	Arg	QG	1.551
25	Arg	QD	3.101
26	Leu	N	119.870
26	Leu	HN	7.833
26	Leu	CA	55.950
26	Leu	HA	4.243
26	Leu	CB	42.520
26	Leu	HB2	1.522
26	Leu	HB3	1.650
26	Leu	HG	1.584
26	Leu	QD1	0.844
26	Leu	QD2	0.877
27	Asn	N	117.025
27	Asn	HN	8.078
27	Asn	CA	53.950
27	Asn	HA	4.481
27	Asn	CB	38.480
27	Asn	HB2	2.447
27	Asn	HB3	2.652
27	Asn	ND2	112.700
27	Asn	HD21	7.248
27	Asn	HD22	6.720
28	Leu	N	120.260
28	Leu	HN	7.976
28	Leu	CA	55.130
28	Leu	HA	4.362

28	Leu	CB	42.620
28	Leu	HB2	1.470
28	Leu	HB3	1.574
28	Leu	HG	1.471
28	Leu	QQD	0.725
29	Gly	N	108.780
29	Gly	HN	8.050
29	Gly	CA	44.943
29	Gly	HA1	3.976
29	Gly	HA2	4.245
30	Pro	CA	64.070
30	Pro	HA	4.369
30	Pro	CB	32.140
30	Pro	HB2	1.813
30	Pro	HB3	2.214
30	Pro	QG	1.962
30	Pro	HD2	3.586
30	Pro	HD3	3.685
31	Ala	N	122.230
31	Ala	HN	8.308
31	Ala	CA	52.800
31	Ala	HA	4.174
31	Ala	QB	1.248
31	Ala	CB	18.700
32	Trp	N	119.100
32	Trp	HN	7.720
32	Trp	CA	57.270
32	Trp	HA	4.562
32	Trp	CB	29.710
32	Trp	HB2	3.213
32	Trp	HB3	3.337
32	Trp	NE1	129.567
32	Trp	HD1	7.224
32	Trp	HE3	7.574
32	Trp	HE1	10.115
32	Trp	HZ3	7.116
32	Trp	HZ2	7.470
33	Gly	N	110.630
33	Gly	HN	8.105
33	Gly	CA	45.809
33	Gly	HA1	3.753
33	Gly	HA2	3.840
34	Gly	N	107.760
34	Gly	HN	7.382
34	Gly	CA	45.137
34	Gly	QA	3.740
35	Arg	N	119.120
35	Arg	HN	7.964
35	Arg	CA	55.930
35	Arg	HA	4.470
35	Arg	CB	31.500
35	Arg	HB2	1.763

35	Arg	HB3	1.899
35	Arg	QG	1.588
35	Arg	QD	3.125
36	Cys	N	125.123
36	Cys	HN	8.212
36	Cys	CA	58.710
36	Cys	HA	4.564
36	Cys	CB	43.090
36	Cys	HB2	3.040
36	Cys	HB3	3.254

Table S2. ^1H chemical shifts of the major *trans* form of SP in TFE/water (1:1) (pH 5.0, 300 K).

Residue	NH	H-C(α)	H-C(β)	Others
Trp ¹	-	4.24	3.19, 3.29	H(ϵ^1) 8.92; H(δ^1) 6.85; H(ϵ^3) 7.51; H(ζ^3) 6.90; H(η^2) 7.15; H(ζ^2) 7.40
Glu ²	7.82	4.44	1.86, 1.91	CH ₂ (γ) 2.04, 2.04
Trp ³	8.14	4.66	2.05, 2.34	H(ϵ^1) 10.04; H(δ^1) 7.36; H(ϵ^3) 7.65; H(ζ^3) 7.42; H(η^2) 7.41; H(ζ^2) 7.63
Pro ⁴	-	4.31	2.05, 2.42	CH ₂ (γ) 2.14, 2.14; CH ₂ (δ) 3.41, 3.91
Trp ⁵	6.77	4.83	3.45, 3.54	H(ϵ^1) 9.93; H(δ^1) 7.22; H(ϵ^3) 7.65; H(ζ^3) 7.30; H(η^2) 7.09; H(ζ^2) 7.00
Asn ⁶	7.89	5.05	2.84, 3.03	NH(ϵ) 7.73; NH(ζ) 7.01
Arg ⁷	7.82	4.45	1.89, 2.00	CH ₂ (γ) 1.71, 1.77; CH ₂ (δ) 3.27, 3.27; NH(ϵ) 7.18; NH ₂ (η) -, -
Lys ⁸	8.29	4.73	1.88, 1.98	CH ₂ (γ) 1.62, 1.62; CH ₂ (δ) 1.87, 1.87; CH ₂ (ϵ) 3.16, 3.16; NH ₃ ⁺ (ζ) -
Hyp ⁹	-	4.67	2.17, 2.51	CH(γ) 4.55; CH ₂ (δ) 3.89, 4.00
Thr ¹⁰	8.32	4.48	4.30	CH ₃ (γ) 1.33
Lys ¹¹	8.16	4.46	1.79, 1.88	CH ₂ (γ) 1.45, 1.49; CH ₂ (δ) 1.79, 1.79; CH ₂ (ϵ) 3.11, 3.11; NH ₃ ⁺ (ζ) -
Phe ¹²	8.13	5.05	3.09, 3.26	H(δ) 7.42, 7.42; H(ϵ) 7.47, 7.47; H(ζ) -
Hyp ¹³	-	4.72	2.20, 2.42	CH(γ) 4.67; CH ₂ (δ) 3.69, 3.95
Ile ¹⁴	8.11	4.66	2.02	CH ₂ (γ') 1.36, 1.74; CH ₃ (γ') 1.16; CH ₃ (δ') 1.06
Hyp ¹⁵	-	4.75	2.22, 2.50	CH(γ) -; CH ₂ (δ) 3.94, 4.11
Ser ¹⁶	8.50	4.95	4.03, 4.03	-
Hyp ¹⁷	-		2.18, 2.50	CH(γ) 4.69; CH ₂ (δ) 4.01, 4.01
Asn ¹⁸	8.81	5.15	3.02, 3.15	NH(ϵ) 7.86; NH(ζ) 6.98
Hyp ¹⁹	-	4.66	2.21, 2.60	CH(γ) 4.61; CH ₂ (δ) 4.10, 4.10
Arg ²⁰	8.32	4.29	1.97, 2.09	CH ₂ (γ) 1.87, 1.87; CH ₂ (δ) 3.34, 3.34; NH(ϵ) 7.38; NH ₂ (η) -, -
Asp ²¹	8.11	4.66	2.96, 3.03	-
Lys ²²	7.91	4.16	1.78, 1.97	CH ₂ (γ) 1.46, 1.55; CH ₂ (δ) 1.79, 1.79; CH ₂ (ϵ) 3.03, 3.03; NH ₃ ⁺ (ζ) -
Trp ²³	8.17	4.44	3.46, 3.61	H(ϵ^1) 9.99; H(δ^1) 7.31; H(ϵ^3) 7.69; H(ζ^3) 7.22; H(η^2) 7.31; H(ζ^2) 7.55
Cys ²⁴	8.62	4.38	3.36, 3.36	
Arg ²⁵	8.18	4.16	1.88, 1.94	CH ₂ (γ) 1.65, 1.74; CH ₂ (δ) 3.16, 3.16; NH(ϵ) 7.38; NH ₂ (η) -, -
Leu ²⁶	8.06	4.29	1.60, 1.80	CH(γ) 1.81; CH ₃ (δ') 0.98; CH ₃ (δ'') 0.98
Asn ²⁷	7.82	4.60	2.28, 2.51	NH(ϵ) 6.79; NH(ζ) 5.84
Leu ²⁸	8.09	4.52	1.53, 1.63	CH(γ) 1.47; CH ₃ (δ') 0.85; CH ₃ (δ'') 0.85
Gly ²⁹	7.93	4.02, 4.42	-	-
Pro ³⁰	-	4.51	2.02, 2.39	CH ₂ (γ) 2.13, 2.13; CH ₂ (δ) 3.73, 3.92
Ala ³¹	8.28	4.45	1.48	

Trp ³²	7.80	4.64	3.39, 3.56	H(ϵ^1) 10.00; H(δ^1) 7.40; H(ϵ^3) 7.72; H(ζ^3) 7.26; H(η^2) 7.34; H(ζ^2) 7.58
Gly ³³	8.11	3.79, 3.96	-	-
Gly ³⁴	7.43	3.80, 3.95	-	-
Arg ³⁵	7.90	4.64	1.96, 2.07	CH ₂ (γ) 1.78, 1.78; CH ₂ (δ) 3.27, 3.27; NH(ϵ) 7.22; NH ₂ (η) -, -
Cys ³⁶	8.17		3.29, 3.46	-

Table S3. NOE upper distance limits in water.

INTRARESIDUAL NOE UPPER-DISTANCE LIMITS

1	TRP	HA	1	TRP	HD1	5.50
1	TRP	HA	1	TRP	HE3	5.50
2	GLU-	HA	2	GLU-	HB2	2.55
2	GLU-	HA	2	GLU-	HB3	2.55
3	TRP	HN	3	TRP	HD1	5.22
3	TRP	HA	3	TRP	HD1	5.50
3	TRP	HA	3	TRP	HE3	4.38
5	TRP	HA	5	TRP	HD1	4.48
5	TRP	HA	5	TRP	HE3	4.88
7	ARG+	HN	7	ARG+	HB2	3.67
7	ARG+	HN	7	ARG+	HB3	3.67
7	ARG+	HN	7	ARG+	QB	3.36
7	ARG+	HN	7	ARG+	HG2	4.97
7	ARG+	HN	7	ARG+	HG3	4.97
7	ARG+	HN	7	ARG+	QG	4.23
7	ARG+	HA	7	ARG+	HG2	3.30
7	ARG+	HA	7	ARG+	HG3	3.30
7	ARG+	HA	7	ARG+	QD	4.67
8	LYS+	HN	8	LYS+	HB2	3.24
8	LYS+	HN	8	LYS+	HB3	3.24
8	LYS+	HN	8	LYS+	QG	6.38
8	LYS+	HA	8	LYS+	HB2	2.90
8	LYS+	HA	8	LYS+	HB3	2.90
8	LYS+	HA	8	LYS+	QB	2.68
10	THR	HN	10	THR	HB	3.42
11	LYS+	HN	11	LYS+	HA	2.90
11	LYS+	HN	11	LYS+	HB2	3.30
11	LYS+	HN	11	LYS+	HB3	3.30
11	LYS+	HN	11	LYS+	QB	2.89
11	LYS+	HN	11	LYS+	HG2	5.50
11	LYS+	HN	11	LYS+	HG3	5.50
11	LYS+	HA	11	LYS+	QG	3.82
12	PHE	HN	12	PHE	HB2	3.73
12	PHE	HN	12	PHE	HB3	3.73
12	PHE	HN	12	PHE	QB	3.38
14	ILE	HN	14	ILE	HA	2.90
14	ILE	HN	14	ILE	HB	3.14
14	ILE	HN	14	ILE	HG12	4.45
14	ILE	HN	14	ILE	HG13	4.45
14	ILE	HN	14	ILE	QD1	6.53
14	ILE	HA	14	ILE	HB	2.90
18	ASN	HN	18	ASN	HB2	3.67
18	ASN	HN	18	ASN	HB3	3.67
18	ASN	HN	18	ASN	QB	3.50
18	ASN	HA	18	ASN	HB2	3.05
18	ASN	HA	18	ASN	HB3	3.05
20	ARG+	HN	20	ARG+	HB2	3.36
20	ARG+	HN	20	ARG+	HB3	3.36

20	ARG+	HN	20	ARG+	QB	3.18
20	ARG+	HN	20	ARG+	QG	5.73
20	ARG+	HA	20	ARG+	QD	6.38
21	ASP-	HN	21	ASP-	HA	2.77
22	LYS+	HN	22	LYS+	QG	5.76
23	TRP	HN	23	TRP	HB2	3.11
23	TRP	HN	23	TRP	HB3	3.11
23	TRP	HN	23	TRP	QB	2.92
23	TRP	HN	23	TRP	HD1	4.88
23	TRP	HN	23	TRP	HE3	5.50
23	TRP	HA	23	TRP	HD1	4.29
23	TRP	HA	23	TRP	HE3	4.45
23	TRP	HB2	23	TRP	HE3	3.79
23	TRP	HB3	23	TRP	HE3	3.79
23	TRP	QB	23	TRP	HE3	3.63
25	ARG+	HN	25	ARG+	HA	2.74
25	ARG+	HN	25	ARG+	HB2	2.99
25	ARG+	HN	25	ARG+	HB3	2.90
25	ARG+	HN	25	ARG+	QG	5.54
25	ARG+	HA	25	ARG+	HB2	2.65
26	LEU	HN	26	LEU	HB2	3.67
26	LEU	HN	26	LEU	HB3	3.67
26	LEU	HN	26	LEU	QB	3.42
26	LEU	HN	26	LEU	HG	4.35
26	LEU	HN	26	LEU	QD1	6.53
26	LEU	HN	26	LEU	QD2	6.53
26	LEU	HA	26	LEU	QD1	5.50
26	LEU	HA	26	LEU	QD2	5.50
27	ASN	HN	27	ASN	HA	2.86
27	ASN	HN	27	ASN	HB2	3.45
27	ASN	HN	27	ASN	HB3	3.45
28	LEU	HN	28	LEU	HB2	3.14
28	LEU	HN	28	LEU	HB3	2.83
28	LEU	HN	28	LEU	QQD	7.63
28	LEU	HA	28	LEU	HB2	2.93
28	LEU	HA	28	LEU	HB3	2.96
29	GLY	HN	29	GLY	HA2	2.93
29	GLY	HN	29	GLY	HA1	2.93
32	TRP	HN	32	TRP	HB2	3.42
32	TRP	HN	32	TRP	HB3	3.42
32	TRP	HA	32	TRP	HD1	4.32
32	TRP	HA	32	TRP	HE3	4.01
32	TRP	HB2	32	TRP	HE3	3.95
32	TRP	HB3	32	TRP	HE3	3.95
35	ARG+	HN	35	ARG+	HA	2.86
35	ARG+	HN	35	ARG+	HB2	3.83
35	ARG+	HN	35	ARG+	HB3	3.83
35	ARG+	HN	35	ARG+	QB	3.38
35	ARG+	HN	35	ARG+	QG	5.26
35	ARG+	HA	35	ARG+	QD	4.27
36	CYSS	HN	36	CYSS	HB3	3.30
36	CYSS	HN	36	CYSS	HB2	3.30

36 CYSS HN 36 CYSS QB 3.10
 SUM = 101

SEQUENTIAL NOE UPPER-DISTANCE LIMITS

2	GLU-	HA	3	TRP	HN	3.14
3	TRP	HA	4	PRO	HD2	3.05
3	TRP	HA	4	PRO	HD3	3.05
3	TRP	HA	4	PRO	QD	2.59
3	TRP	QB	4	PRO	HD2	6.31
3	TRP	QB	4	PRO	HD3	6.31
3	TRP	QB	4	PRO	QD	6.06
3	TRP	HE3	4	PRO	HD2	5.19
3	TRP	HE3	4	PRO	HD3	5.19
3	TRP	HE3	4	PRO	QD	4.76
4	PRO	HA	5	TRP	HN	3.14
4	PRO	QB	5	TRP	HN	4.48
4	PRO	QD	5	TRP	HN	5.69
5	TRP	HN	6	ASN	HN	3.42
5	TRP	HA	6	ASN	HN	3.30
5	TRP	QB	6	ASN	HN	4.52
5	TRP	HD1	6	ASN	HN	5.31
5	TRP	HE3	6	ASN	HN	5.50
6	ASN	HA	7	ARG+	HN	3.36
7	ARG+	HN	8	LYS+	HN	3.64
7	ARG+	HA	8	LYS+	HN	2.74
8	LYS+	HN	9	HYPR	QD	6.20
8	LYS+	HA	9	HYPR	HD2	3.17
8	LYS+	HA	9	HYPR	HD3	3.17
8	LYS+	HA	9	HYPR	QD	2.83
8	LYS+	HB2	9	HYPR	HD2	5.50
8	LYS+	HB2	9	HYPR	HD3	5.50
8	LYS+	HB3	9	HYPR	HD2	5.50
8	LYS+	HB3	9	HYPR	HD3	5.50
8	LYS+	QG	9	HYPR	HD2	6.38
8	LYS+	QG	9	HYPR	HD3	6.38
9	HYPR	HA	10	THR	HN	2.77
9	HYPR	HB2	10	THR	HN	4.04
9	HYPR	HB3	10	THR	HN	4.04
10	THR	HA	11	LYS+	HN	2.86
10	THR	HB	11	LYS+	HN	3.30
10	THR	QG2	11	LYS+	HN	6.53
11	LYS+	HA	12	PHE	HN	2.65
11	LYS+	HA	12	PHE	QD	7.62
12	PHE	HA	13	HYPR	HD2	3.45
12	PHE	HA	13	HYPR	HD3	3.45
12	PHE	HA	13	HYPR	QD	3.07
12	PHE	HB2	13	HYPR	HG2	5.50
12	PHE	HB2	13	HYPR	HD2	5.38
12	PHE	HB2	13	HYPR	HD3	5.38
12	PHE	HB3	13	HYPR	HG2	5.50

12	PHE	HB3	13	HYPR	HD2	5.38
12	PHE	HB3	13	HYPR	HD3	5.38
12	PHE	QB	13	HYPR	QD	4.39
12	PHE	QD	13	HYPR	HD2	7.62
12	PHE	QD	13	HYPR	HD3	7.62
13	HYPR	HA	14	ILE	HN	2.49
13	HYPR	HB2	14	ILE	HN	4.17
13	HYPR	HB3	14	ILE	HN	4.17
13	HYPR	QB	14	ILE	HN	3.92
14	ILE	HN	15	HYPR	HD2	5.50
14	ILE	HN	15	HYPR	HD3	5.50
14	ILE	HA	15	HYPR	HD2	2.71
14	ILE	HA	15	HYPR	HD3	2.71
14	ILE	HB	15	HYPR	HD2	4.88
14	ILE	HB	15	HYPR	HD3	4.88
14	ILE	HB	15	HYPR	QD	4.12
14	ILE	QG2	15	HYPR	HD2	5.66
14	ILE	QG2	15	HYPR	HD3	5.66
14	ILE	QG2	15	HYPR	QD	5.34
15	HYPR	HA	16	SER	HN	2.59
15	HYPR	HB2	16	SER	HN	3.89
15	HYPR	HB3	16	SER	HN	3.89
15	HYPR	QB	16	SER	HN	3.72
16	SER	HA	17	HYPR	QD	4.69
17	HYPR	HA	18	ASN	HN	2.86
17	HYPR	HB2	18	ASN	HN	4.29
17	HYPR	HB3	18	ASN	HN	4.29
17	HYPR	QB	18	ASN	HN	4.07
18	ASN	HN	19	HYPR	QD	6.39
18	ASN	HA	19	HYPR	QD	3.82
18	ASN	HB2	19	HYPR	QD	5.87
18	ASN	HB3	19	HYPR	QD	5.87
19	HYPR	HA	20	ARG+	HN	2.80
19	HYPR	HB2	20	ARG+	HN	4.26
19	HYPR	HB3	20	ARG+	HN	4.26
19	HYPR	QB	20	ARG+	HN	4.01
20	ARG+	HN	21	ASP-	HN	3.14
20	ARG+	HA	21	ASP-	HN	2.99
21	ASP-	HA	22	LYS+	HN	2.77
21	ASP-	QB	22	LYS+	HN	4.11
22	LYS+	HA	23	TRP	HN	3.17
22	LYS+	HA	23	TRP	HD1	5.50
23	TRP	HN	24	CYSS	HN	5.50
23	TRP	HA	24	CYSS	HN	2.83
23	TRP	HB2	24	CYSS	HN	3.89
23	TRP	HB3	24	CYSS	HN	3.89
23	TRP	QB	24	CYSS	HN	3.71
23	TRP	HD1	24	CYSS	HN	5.50
23	TRP	HD1	24	CYSS	HA	4.63
23	TRP	HE3	24	CYSS	HN	5.50
23	TRP	HE3	24	CYSS	HA	4.57
23	TRP	HE3	24	CYSS	QB	6.38

24	CYSS	HA	25	ARG+	HN	2.80
24	CYSS	QB	25	ARG+	HN	4.36
25	ARG+	HN	26	LEU	HN	5.50
25	ARG+	HA	26	LEU	HN	3.39
25	ARG+	HB2	26	LEU	HN	3.79
25	ARG+	HB3	26	LEU	HN	3.45
26	LEU	HN	27	ASN	HN	4.35
26	LEU	HA	27	ASN	HN	2.90
26	LEU	HB2	27	ASN	HN	3.24
26	LEU	HB3	27	ASN	HN	3.24
26	LEU	QB	27	ASN	HN	2.96
26	LEU	QQD	27	ASN	HN	8.10
28	LEU	HA	29	GLY	HN	2.77
28	LEU	HB2	29	GLY	HN	3.55
28	LEU	HB3	29	GLY	HN	3.11
28	LEU	QQD	29	GLY	HN	7.63
29	GLY	HN	30	PRO	QD	5.74
29	GLY	HA2	30	PRO	HD2	3.73
29	GLY	HA2	30	PRO	HD3	3.73
29	GLY	HA1	30	PRO	HD2	3.73
29	GLY	HA1	30	PRO	HD3	3.73
29	GLY	QA	30	PRO	QD	3.00
30	PRO	HA	31	ALA	HN	3.02
30	PRO	QB	31	ALA	HN	4.70
30	PRO	HD2	31	ALA	HN	5.50
30	PRO	HD3	31	ALA	HN	5.50
30	PRO	QD	31	ALA	HN	5.35
31	ALA	HN	32	TRP	HN	3.64
31	ALA	HA	32	TRP	HN	3.36
31	ALA	QB	32	TRP	HD1	8.00
32	TRP	HN	33	GLY	HN	3.98
32	TRP	HA	33	GLY	HN	3.05
32	TRP	HB2	33	GLY	HN	4.11
32	TRP	HB3	33	GLY	HN	4.11
32	TRP	QB	33	GLY	HN	3.89
33	GLY	HN	34	GLY	HN	3.76
33	GLY	HA2	34	GLY	HN	3.14
33	GLY	HA1	34	GLY	HN	3.14
33	GLY	QA	34	GLY	HN	2.84
35	ARG+	HA	36	CYSS	HN	2.68
SUM = 138						

MEDIUM and LONG RANGE NOE UPPER-DISTANCE LIMITS

3	TRP	QB	5	TRP	HN	5.32
3	TRP	QB	5	TRP	HE1	6.38
10	THR	QG2	12	PHE	HN	6.53
10	THR	QG2	12	PHE	QD	8.65
18	ASN	HN	21	ASP-	QB	5.38
18	ASN	HA	20	ARG+	HN	5.50
19	HYPR	QB	21	ASP-	HN	5.70

20	ARG+	HA	23	TRP	HD1	5.50
20	ARG+	HA	23	TRP	HE3	5.50
20	ARG+	HA	23	TRP	HE1	5.50
23	TRP	HB2	28	LEU	QQD	7.63
23	TRP	HB3	28	LEU	QQD	7.63
23	TRP	HE3	28	LEU	QQD	7.63
24	CYSS	HA	26	LEU	HN	3.30
24	CYSS	QB	26	LEU	HN	6.31
24	CYSS	QB	28	LEU	QQD	8.51
24	CYSS	QB	32	TRP	HE3	6.38
28	LEU	HB2	32	TRP	HD1	4.63
28	LEU	HB3	32	TRP	HD1	4.38
28	LEU	QQD	32	TRP	HB2	7.63
28	LEU	QQD	32	TRP	HB3	7.63
28	LEU	QQD	32	TRP	HD1	7.63
28	LEU	QQD	32	TRP	HE3	7.63
28	LEU	QQD	32	TRP	HE1	7.63
28	LEU	QQD	32	TRP	HZ3	7.63
29	GLY	QA	31	ALA	HN	4.55
30	PRO	HA	32	TRP	HN	4.14
SUM = 27						

Table S4. NOE upper distance limits in TFE/water.

INTRARESIDUAL NOE UPPER-DISTANCE LIMITS

1	TRP	HA	1	TRP	HD1	6.00
1	TRP	HA	1	TRP	HE3	5.39
1	TRP	HB2	1	TRP	HD1	3.70
1	TRP	HB3	1	TRP	HD1	3.70
1	TRP	QB	1	TRP	HD1	3.54
2	GLU-	HN	2	GLU-	HB2	3.55
2	GLU-	HN	2	GLU-	HB3	3.55
2	GLU-	HN	2	GLU-	QG	6.70
2	GLU-	HA	2	GLU-	HB2	2.76
2	GLU-	HA	2	GLU-	HB3	2.76
2	GLU-	HA	2	GLU-	QB	2.49
3	TRP	HN	3	TRP	QB	3.69
3	TRP	HN	3	TRP	HD1	4.78
3	TRP	HA	3	TRP	HD1	6.00
3	TRP	HA	3	TRP	HE3	4.09
5	TRP	HN	5	TRP	HB2	4.02
5	TRP	HN	5	TRP	HB3	4.02
5	TRP	HN	5	TRP	QB	3.72
5	TRP	HN	5	TRP	HE3	4.52
5	TRP	HA	5	TRP	HD1	6.00
5	TRP	HA	5	TRP	HE3	6.00
6	ASN	HN	6	ASN	HB2	3.62
6	ASN	HN	6	ASN	HB3	3.62
6	ASN	HN	6	ASN	QB	3.42

6	ASN	HA	6	ASN	HD21	6.00
6	ASN	HA	6	ASN	HD22	6.00
7	ARG+	HN	7	ARG+	HB2	3.52
7	ARG+	HN	7	ARG+	HB3	3.52
7	ARG+	HN	7	ARG+	HG2	5.60
7	ARG+	HN	7	ARG+	HG3	5.60
7	ARG+	HN	7	ARG+	QG	5.00
7	ARG+	HA	7	ARG+	HB2	2.94
7	ARG+	HA	7	ARG+	HB3	2.94
7	ARG+	HA	7	ARG+	HG2	3.55
7	ARG+	HA	7	ARG+	HG3	3.55
7	ARG+	HA	7	ARG+	QG	3.26
7	ARG+	HB2	7	ARG+	HE	6.00
7	ARG+	HB3	7	ARG+	HE	6.00
8	LYS+	HN	8	LYS+	HB2	3.16
8	LYS+	HN	8	LYS+	HB3	3.16
8	LYS+	HN	8	LYS+	QB	2.92
8	LYS+	HN	8	LYS+	QG	6.26
10	THR	HN	10	THR	HB	3.16
11	LYS+	HN	11	LYS+	HA	2.76
11	LYS+	HN	11	LYS+	HB2	3.12
11	LYS+	HN	11	LYS+	HB3	3.12
11	LYS+	HN	11	LYS+	QB	2.88
11	LYS+	HN	11	LYS+	HG2	4.70
11	LYS+	HN	11	LYS+	HG3	4.70
11	LYS+	HN	11	LYS+	QG	4.06
11	LYS+	HA	11	LYS+	QG	3.71
12	PHE	HN	12	PHE	HB2	3.37
12	PHE	HN	12	PHE	HB3	3.37
12	PHE	HN	12	PHE	QB	3.02
14	ILE	HN	14	ILE	HB	3.00
14	ILE	HN	14	ILE	HG12	4.45
14	ILE	HN	14	ILE	HG13	4.45
14	ILE	HN	14	ILE	QG1	3.76
14	ILE	HN	14	ILE	QD1	6.74
18	ASN	HN	18	ASN	HB2	3.59
18	ASN	HN	18	ASN	HB3	3.59
18	ASN	HN	18	ASN	QB	3.40
20	ARG+	HN	20	ARG+	HB2	3.30
20	ARG+	HN	20	ARG+	HB3	3.30
20	ARG+	HN	20	ARG+	QB	3.07
20	ARG+	HN	20	ARG+	QG	5.65
21	ASP-	HN	21	ASP-	HB2	3.48
21	ASP-	HN	21	ASP-	HB3	3.48
21	ASP-	HN	21	ASP-	QB	3.19
22	LYS+	HN	22	LYS+	HB2	3.23
22	LYS+	HN	22	LYS+	HB3	3.23
22	LYS+	HN	22	LYS+	QB	2.96
22	LYS+	HN	22	LYS+	HG2	4.88
22	LYS+	HN	22	LYS+	HG3	4.88
22	LYS+	HN	22	LYS+	QG	4.64
22	LYS+	HA	22	LYS+	HG2	3.84

22	LYS+	HA	22	LYS+	HG3	3.84
22	LYS+	HA	22	LYS+	QG	3.68
22	LYS+	QB	22	LYS+	QE	6.64
23	TRP	HN	23	TRP	HB2	3.41
23	TRP	HN	23	TRP	HB3	3.41
23	TRP	HN	23	TRP	QB	3.13
23	TRP	HN	23	TRP	HD1	4.67
23	TRP	HN	23	TRP	HE3	5.00
23	TRP	HA	23	TRP	HD1	4.56
23	TRP	HA	23	TRP	HE3	4.13
23	TRP	HB2	23	TRP	HE3	4.02
23	TRP	HB3	23	TRP	HE3	4.02
23	TRP	QB	23	TRP	HE1	5.18
25	ARG+	HN	25	ARG+	HB2	3.30
25	ARG+	HN	25	ARG+	HB3	3.30
25	ARG+	HA	25	ARG+	HB2	2.90
25	ARG+	HA	25	ARG+	HB3	2.90
25	ARG+	HA	25	ARG+	HG2	3.52
25	ARG+	HA	25	ARG+	HG3	3.52
25	ARG+	HA	25	ARG+	QG	3.12
25	ARG+	HA	25	ARG+	HE	5.89
25	ARG+	QG	25	ARG+	HE	3.85
26	LEU	HN	26	LEU	HB2	2.98
26	LEU	HN	26	LEU	HB3	3.23
26	LEU	HN	26	LEU	QQD	7.56
26	LEU	HA	26	LEU	HB2	2.65
26	LEU	HA	26	LEU	HB3	2.98
27	ASN	HN	27	ASN	HB2	3.59
27	ASN	HN	27	ASN	HB3	3.59
27	ASN	HN	27	ASN	QB	3.39
27	ASN	HN	27	ASN	QD2	5.32
28	LEU	HN	28	LEU	QQD	8.02
31	ALA	HN	31	ALA	HA	2.83
32	TRP	HN	32	TRP	HB2	3.19
32	TRP	HN	32	TRP	HB3	3.19
32	TRP	HN	32	TRP	HD1	4.60
32	TRP	HA	32	TRP	HD1	5.10
32	TRP	HA	32	TRP	HE3	4.45
32	TRP	HB2	32	TRP	HE3	3.88
32	TRP	HB3	32	TRP	HE3	3.88
32	TRP	QB	32	TRP	HE3	3.69
35	ARG+	HN	35	ARG+	HB2	3.41
35	ARG+	HN	35	ARG+	HB3	3.41
35	ARG+	HN	35	ARG+	QB	3.13
35	ARG+	HN	35	ARG+	QG	4.68
35	ARG+	HB2	35	ARG+	HE	5.53
35	ARG+	HB3	35	ARG+	HE	5.53
35	ARG+	QB	35	ARG+	HE	5.37
36	CYSS	HN	36	CYSS	HB3	3.23
36	CYSS	HN	36	CYSS	HB2	3.23
36	CYSS	HN	36	CYSS	QB	3.00

SUM = 127

SEQUENTIAL NOE UPPER-DISTANCE LIMITS

1	TRP	HA	2	GLU-	HN	3.41
1	TRP	HB2	2	GLU-	HN	4.27
1	TRP	HB3	2	GLU-	HN	4.27
1	TRP	QB	2	GLU-	HN	4.07
1	TRP	HE3	2	GLU-	HA	6.00
2	GLU-	HA	3	TRP	HN	3.05
3	TRP	HA	4	PRO	HD2	3.30
3	TRP	HA	4	PRO	HD3	3.30
3	TRP	HA	4	PRO	QD	2.96
3	TRP	HB2	4	PRO	HD2	6.99
3	TRP	HB2	4	PRO	HD3	6.99
3	TRP	HB3	4	PRO	HD2	6.99
3	TRP	HB3	4	PRO	HD3	6.99
3	TRP	QB	4	PRO	QD	6.02
4	PRO	HA	5	TRP	HN	3.59
4	PRO	HB2	5	TRP	HN	4.67
4	PRO	HB3	5	TRP	HN	4.67
4	PRO	QB	5	TRP	HN	4.05
4	PRO	QB	5	TRP	HZ3	6.02
4	PRO	QG	5	TRP	HE3	6.71
4	PRO	QG	5	TRP	HZ3	6.89
4	PRO	HD2	5	TRP	HE3	4.38
4	PRO	HD3	5	TRP	HE3	4.38
4	PRO	QD	5	TRP	HN	5.05
4	PRO	QD	5	TRP	HE3	4.09
5	TRP	HN	6	ASN	HN	3.62
5	TRP	QB	6	ASN	HN	4.61
5	TRP	HD1	6	ASN	HN	6.00
5	TRP	HD1	6	ASN	HA	6.00
5	TRP	HE3	6	ASN	HN	4.67
6	ASN	HA	7	ARG+	HN	3.30
6	ASN	HB2	7	ARG+	HN	4.24
6	ASN	HB3	7	ARG+	HN	4.24
6	ASN	QB	7	ARG+	HN	3.88
7	ARG+	HN	8	LYS+	HN	3.41
7	ARG+	HA	8	LYS+	HN	2.98
8	LYS+	HN	9	HYPR	QD	6.13
8	LYS+	HA	9	HYPR	HD2	4.13
8	LYS+	HA	9	HYPR	HD3	4.13
8	LYS+	HA	9	HYPR	QD	3.80
8	LYS+	HB2	9	HYPR	HD2	6.00
8	LYS+	HB2	9	HYPR	HD3	6.00
8	LYS+	HB3	9	HYPR	HD2	6.00
8	LYS+	HB3	9	HYPR	HD3	6.00
8	LYS+	QB	9	HYPR	QD	4.97
8	LYS+	QG	9	HYPR	HD2	6.88
8	LYS+	QG	9	HYPR	HD3	6.88
9	HYPR	HB2	10	THR	HN	4.38

9	HYPR	HB3	10	THR	HN	4.38
9	HYPR	QB	10	THR	HN	4.16
10	THR	HA	11	LYS+	HN	2.83
10	THR	HB	11	LYS+	HN	3.08
10	THR	QG2	11	LYS+	HN	5.80
11	LYS+	HA	12	PHE	HN	2.94
12	PHE	HA	13	HYPR	HD2	3.30
12	PHE	HA	13	HYPR	HD3	3.30
12	PHE	HA	13	HYPR	QD	2.96
12	PHE	HB2	13	HYPR	HD2	5.14
12	PHE	HB2	13	HYPR	HD3	5.14
12	PHE	HB3	13	HYPR	HD2	5.14
12	PHE	HB3	13	HYPR	HD3	5.14
12	PHE	QB	13	HYPR	QD	4.25
12	PHE	QD	13	HYPR	HD2	8.12
12	PHE	QD	13	HYPR	HD3	8.12
12	PHE	QD	13	HYPR	QD	7.66
13	HYPR	HB2	14	ILE	HN	3.95
13	HYPR	HB3	14	ILE	HN	3.95
13	HYPR	QB	14	ILE	HN	3.76
14	ILE	HA	15	HYPR	HD2	3.59
14	ILE	HA	15	HYPR	HD3	3.59
14	ILE	HA	15	HYPR	QD	3.08
14	ILE	HB	15	HYPR	HD2	4.06
14	ILE	HB	15	HYPR	HD3	4.06
14	ILE	HB	15	HYPR	QD	3.23
14	ILE	QG2	15	HYPR	HD2	5.91
14	ILE	QG2	15	HYPR	HD3	5.91
14	ILE	QG2	15	HYPR	QD	5.52
14	ILE	QG1	15	HYPR	QD	7.77
14	ILE	QD1	15	HYPR	HD2	7.03
14	ILE	QD1	15	HYPR	HD3	7.03
15	HYPR	HB2	16	SER	HN	4.34
15	HYPR	HB3	16	SER	HN	4.34
15	HYPR	QB	16	SER	HN	4.16
16	SER	HA	17	HYPR	QD	3.98
17	HYPR	HB2	18	ASN	HN	4.63
17	HYPR	HB3	18	ASN	HN	4.63
17	HYPR	QB	18	ASN	HN	4.42
18	ASN	HN	19	HYPR	QD	6.89
18	ASN	HA	19	HYPR	QD	4.16
18	ASN	HB2	19	HYPR	QD	6.14
18	ASN	HB3	19	HYPR	QD	6.14
18	ASN	QB	19	HYPR	QD	5.57
19	HYPR	HB2	20	ARG+	HN	4.38
19	HYPR	HB3	20	ARG+	HN	4.38
19	HYPR	QB	20	ARG+	HN	4.06
20	ARG+	HN	21	ASP-	HN	2.40
20	ARG+	HA	21	ASP-	HN	3.12
20	ARG+	QB	21	ASP-	HN	4.22
21	ASP-	HN	22	LYS+	HN	2.44
21	ASP-	HA	22	LYS+	HN	3.48

21	ASP-	HB2	22	LYS+	HN	3.80
21	ASP-	HB3	22	LYS+	HN	3.80
21	ASP-	QB	22	LYS+	HN	3.63
22	LYS+	HN	23	TRP	HN	3.20
22	LYS+	HA	23	TRP	HN	3.50
22	LYS+	HB2	23	TRP	HB2	6.81
22	LYS+	HB2	23	TRP	HB3	6.81
22	LYS+	HB3	23	TRP	HB2	6.81
22	LYS+	HB3	23	TRP	HB3	6.81
22	LYS+	QB	23	TRP	QB	5.18
23	TRP	HN	24	CYSS	HN	3.59
23	TRP	HB2	24	CYSS	HN	3.70
23	TRP	HB3	24	CYSS	HN	3.70
23	TRP	QB	24	CYSS	HN	3.54
23	TRP	HD1	24	CYSS	HN	6.00
23	TRP	HE3	24	CYSS	HN	5.50
23	TRP	HE3	24	CYSS	HA	4.45
23	TRP	HE3	24	CYSS	QB	6.84
24	CYSS	HN	25	ARG+	HN	3.48
24	CYSS	HA	25	ARG+	HN	3.12
24	CYSS	QB	25	ARG+	HN	4.46
25	ARG+	HN	26	LEU	HN	3.00
25	ARG+	HA	26	LEU	HN	3.60
25	ARG+	HB2	26	LEU	HN	3.52
25	ARG+	HB3	26	LEU	HN	3.52
25	ARG+	QB	26	LEU	HN	3.26
26	LEU	HN	27	ASN	HN	3.20
26	LEU	HA	27	ASN	HN	3.50
26	LEU	HB2	27	ASN	HN	3.66
26	LEU	HB2	27	ASN	HD21	6.00
26	LEU	HB2	27	ASN	HD22	6.00
26	LEU	HB2	27	ASN	QD2	5.16
26	LEU	HB3	27	ASN	HN	3.44
26	LEU	HB3	27	ASN	HD21	6.00
26	LEU	HB3	27	ASN	HD22	6.00
26	LEU	QQD	27	ASN	HN	8.13
27	ASN	HN	28	LEU	QQD	8.13
27	ASN	HA	28	LEU	HN	3.19
27	ASN	HB2	28	LEU	HN	4.30
27	ASN	HB2	28	LEU	QQD	8.13
27	ASN	HB3	28	LEU	HN	4.30
27	ASN	HB3	28	LEU	QQD	8.13
28	LEU	HA	29	GLY	HN	3.37
28	LEU	HB2	29	GLY	HN	3.95
28	LEU	HB3	29	GLY	HN	3.95
28	LEU	QB	29	GLY	HN	3.50
28	LEU	QQD	29	GLY	HN	8.13
29	GLY	HN	30	PRO	HD2	4.45
29	GLY	HN	30	PRO	HD3	4.45
29	GLY	HA2	30	PRO	HD2	5.23
29	GLY	HA2	30	PRO	HD3	5.23
29	GLY	HA1	30	PRO	HD2	5.23

29	GLY	HA1	30	PRO	HD3	5.23
29	GLY	QA	30	PRO	QD	4.19
30	PRO	HA	31	ALA	HN	3.70
30	PRO	HB2	31	ALA	HN	4.42
30	PRO	HB3	31	ALA	HN	4.42
30	PRO	QB	31	ALA	HN	3.88
30	PRO	HD2	31	ALA	HN	5.28
30	PRO	HD3	31	ALA	HN	5.28
30	PRO	QD	31	ALA	HN	4.96
31	ALA	HN	32	TRP	HN	3.37
31	ALA	HN	32	TRP	HD1	6.00
31	ALA	HA	32	TRP	HN	3.62
31	ALA	HA	32	TRP	HD1	5.80
31	ALA	QB	32	TRP	HD1	6.60
32	TRP	HN	33	GLY	HN	3.20
32	TRP	HA	33	GLY	HN	3.50
32	TRP	HB2	33	GLY	HN	3.41
32	TRP	HB3	33	GLY	HN	3.41
32	TRP	HD1	33	GLY	HN	6.00
32	TRP	HE3	33	GLY	HA2	6.00
32	TRP	HE3	33	GLY	HA1	6.00
32	TRP	HE3	33	GLY	QA	5.56
33	GLY	HN	34	GLY	HN	3.48
33	GLY	HA2	34	GLY	HN	3.23
33	GLY	HA1	34	GLY	HN	3.23
33	GLY	QA	34	GLY	HN	3.03
34	GLY	HN	35	ARG+	HN	3.84
34	GLY	HA2	35	ARG+	HN	3.34
34	GLY	HA1	35	ARG+	HN	3.34
35	ARG+	HA	36	CYSS	HN	2.98
SUM = 182						

MEDIUM and LONG RANGE NOE UPPER-DISTANCE LIMITS

3	TRP	HN	6	ASN	QB	4.86
3	TRP	HB2	5	TRP	HE1	6.00
3	TRP	HB3	5	TRP	HE1	6.00
3	TRP	QB	5	TRP	HN	5.22
3	TRP	QB	5	TRP	HD1	6.88
3	TRP	QB	5	TRP	HZ2	6.88
3	TRP	QB	6	ASN	HN	5.01
3	TRP	HD1	6	ASN	QD2	4.78
10	THR	QG2	12	PHE	HN	6.96
10	THR	QG2	12	PHE	QD	9.15
10	THR	QG2	12	PHE	QE	9.15
18	ASN	HA	20	ARG+	HN	4.60
18	ASN	HB2	20	ARG+	HN	4.31
18	ASN	HB3	20	ARG+	HN	4.31
18	ASN	QB	20	ARG+	HN	4.07
20	ARG+	HA	23	TRP	HD1	5.68
20	ARG+	HA	23	TRP	HE3	4.92

20	ARG+	HA	24	CYSS	HN	4.38
21	ASP-	HA	24	CYSS	HN	4.70
22	LYS+	HA	24	CYSS	HN	4.13
23	TRP	HA	27	ASN	HD21	5.50
23	TRP	HA	27	ASN	HD22	5.50
23	TRP	HA	27	ASN	QD2	5.32
23	TRP	HD1	26	LEU	QQD	8.13
23	TRP	HD1	28	LEU	QQD	8.13
23	TRP	HE3	26	LEU	QQD	8.13
23	TRP	HE3	28	LEU	QQD	8.13
23	TRP	HZ3	26	LEU	QQD	8.13
23	TRP	HZ3	28	LEU	QQD	8.13
23	TRP	HH2	28	LEU	QQD	8.13
24	CYSS	QB	26	LEU	HN	6.41
24	CYSS	QB	28	LEU	QQD	9.01
26	LEU	HN	28	LEU	QQD	8.13
26	LEU	QQD	28	LEU	HN	8.13
28	LEU	HA	32	TRP	HD1	5.32
28	LEU	QB	32	TRP	HD1	6.63
28	LEU	QQD	32	TRP	HN	8.13
28	LEU	QQD	32	TRP	HB2	8.13
28	LEU	QQD	32	TRP	HB3	8.13
28	LEU	QQD	32	TRP	QB	7.90
28	LEU	QQD	32	TRP	HD1	8.13
28	LEU	QQD	32	TRP	HE3	8.13
28	LEU	QQD	32	TRP	HE1	8.13
28	LEU	QQD	32	TRP	HZ2	8.13
28	LEU	QQD	36	CYSS	HN	8.13
28	LEU	QQD	36	CYSS	HB3	8.13
28	LEU	QQD	36	CYSS	HB2	8.13
28	LEU	QQD	36	CYSS	QB	7.99
29	GLY	QA	31	ALA	HN	5.08
30	PRO	HA	32	TRP	HN	3.70
32	TRP	HE3	36	CYSS	QB	6.48

SUM = 51

Figure-S1. ^{15}N -Chemical shift deviations from random coil for residues 1-36 in SP ($\Delta\delta = \delta^{\text{observed}} - \delta^{\text{random}}$) in water solution. The asterik shows the positions of hydroxy-proline (Hyp) in the sequence.

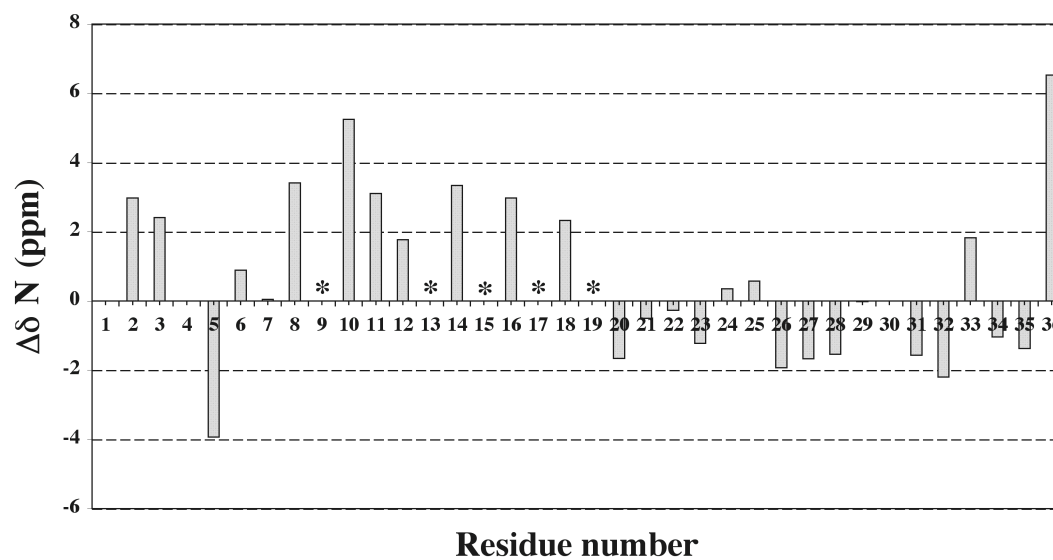


Figure-S2. Results from the MD simulation of SP with time-averaged NOE-derived distance restraints. The ϕ (black lines), ψ (gray lines) backbone torsion angles of each residue in SP, from Glu² to Arg³⁵, as a function of simulation time in ps.

